



Full Article

Analytical Method Validation of Selected Organochlorine Pesticides in Fortified Rice Using Gas Chromatography with Electron Capture Detector

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ABSTRACT

Gas chromatography with electron capture detector (GC-ECD) has been validated for simultaneous determination of selected organochlorine pesticides namely heptachlor, endosulfan, dieldrin, dan *p,p'*-dichlorodiphenyltrichloroethane (*p,p'*-DDT) using aldrin as standard internal. Some parameters for analytical method validation were evaluated, i.e. linearity, precision, sensitivity expressed with limit of detection and limit of quantification, and recovery. GC-ECD was linear for determination of these pesticides with *r* values ≥ 0.99 . The values of LOD obtained were 2.50, 2.97, 1.61, and 2.54 ng/mL for heptachlor, endosulphan, dieldrin, and *p,p'*-DDT, respectively. Furthermore, the relative standard deviation (RSD) values obtained for the assessment of analytical precision were lower than those required by Horwitz. Finally, the used GC-ECD method was accurate for determination of these organochlorine pesticides in fortified rice samples which were indicated by the acceptable recovery percentages.

Keywords: validation, organochlorine pesticides, gas chromatography, electron capture detector, rice

1. Introduction

Currently, Indonesian is one of the countries consuming rice as its food staple. Rice is one of the important human diets as carbohydrate source obtained from the paddy (*Oryza sativa* L). In order to increase the productivity of paddy, the farmers used pesticides. Pesticides are chemical compounds, which are frequently used in modern agriculture practices to keep the crops from different pests and diseases (Guler *et al.*, 2010).

The use of pesticides is important in modern agriculture in order to eliminate the disease as well as to enhance the productivity of paddy (Juraske *et al.*, 2009); however, the application of pesticides in paddy can lead to some drawbacks in the form of pesticide residues remaining in paddy and of course in rice. This can cause the human health risks who consumed rice (Nia *et al.*, 2009), consequently rice containing pesticide residue

can be potential problem (Radwan and Salama, 2006). Therefore, pesticides should be controlled at optimum level due to their relative toxicity to the human health and the environment (Jiang *et al.*, 2009).

In this study, we selected four organochlorine pesticides namely heptachlor, endosulfan, dieldrin, dan *p,p'*-DDT. The chemical structures of these pesticides were shown in Figure 1. Heptachlor is chlorinated dicyclopentadiene pesticide (Fig. 1) which is persistent in environment and can accumulate in food chain. Since 1980's, this pesticide has been banned to be used in agriculture, however, heptachlor is still detected in some food commodities (Kielhorn *et al.*, 2006). Endosulfan is used as an insecticide and acaricide. This compound has a highly controversial agrichemical due to its acute toxicity, potential for bioaccumulation, and its role as an endocrine disruptor (Latif *et al.*, 2011). Furthermore, in some countries, dieldrin and *p,p'*-DDT are restricted to be used in agriculture field (Ritter *et al.*, 1995).

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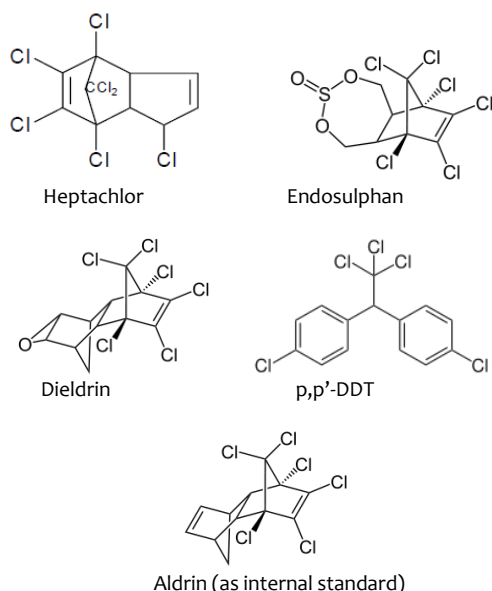


Fig 1. The chemical structures of studied organochlorine pesticides

Several analytical methods have been published for determination of organochlorine pesticides in numerous food matrixes such as high performance liquid chromatography (Trajkovska *et al.*, 2003; Vega-Moreno *et al.*, 2008), liquid chromatography tandem with mass spectrometer known as LC-MS (Paíga *et al.*, 2009), gas chromatography-mass spectrometry (Latif *et al.*, 2011, Yang *et al.*, 2011), gas chromatography using nitrogen-phosphorus detector (Athanasopoulos and Pappas, 2000; Albero *et al.*, 2003; Lambropoulou and Albanis, 2002), flame photometric detector (Zabik *et al.* 2000; Simplicio and Boas, 1999), and electron capture detector (Abou-Arab, 1999; Cai *et al.*, 2005). To our knowledge, there is no available reports related the use of GC in combination with electron capture detector for quantitative analysis of selected organochlorine pesticides. Therefore, in this study, we validated GC-ECD for such analysis and applied the validated method for quantitative analysis of selected organochlorine pesticides in fortified rice samples.

2. Materials and Methods

The organochlorine pesticides of heptachlor, endosulfan, dieldrin, and *p,p'*-DDT as well as internal standard of aldrin were bought from Aldrich, USA; activated carbon, toluene, 2-propanol and sodium sulphate anhydrate were purchased from E. Merck (Darmstadt Germany). Celite 545 was obtained from Gasukuro Kogyo, Japan. The rice was obtained from local market in Yogyakarta, Indonesia.

2.2. Sample preparation

An approximately of 10,0 g rice sample was accurately weighed using analytical balance with sensitivity of 0.1 mg in Erlenmeyer. The sample was added with aldrin as internal standard, 20 mL toluene and 10 mL 2-propanol. The mixture was vigorously blended with *ultra turrax* for 5 min. The mixture obtained was allowed for 5 min to precipitate and subsequently filtered. The filtrate was extracted using 2 x 50 mL

Na₂SO₄ 2 % b/v in water. The aqueous phase was removed and organic phase (toluene) was taken for clean up procedure.

2.2. Clean up

An approximately of 10.0 mL toluene phase was taken and introduced to closed-reaction. After that, 1 gram of adsorbent (celite 545 and active carbon (1:3, w/w)) was added into reaction tube. The mixture was vigorously shaken for 1 min and filtered using Whatman paper No. 1. The supernatant was directly injected (1 µL) to gas chromatograph.

2.3. GC condition

The GC condition was adopted from the previously reported by Anugrahwati (2010) using the same instrument, as below:

| | |
|----------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Column | : Rtx-1301 (fused silica capillary column) from Restex, Bellafante, USA (30 m x 0.25 i.d; film thickness 0,25 µm). the column was maintained isothermally at 230°C |
| Carrier gas | : Helium (1.7 mL/min) |
| Make up gas | : N ₂ / air |
| Detector | : Electron capture detector (300°C) |
| Injector temperature | : 270°C |
| Injection volume | : 1 µl |
| Detector | : ECD (Electron Capture Detector) at 300 °C |

2.4. Analytical method validation

Several analytical figure of merits was evaluated for determining validation criteria, namely linearity, precision, sensitivity expressed as limit of detection and limit of quantification, and accuracy (ICH, 1994).

3. Results and Discussion

Several pesticides have specific properties, namely low-polarity, stable toward thermal, and volatile compounds. Consequently, they can be analyzed using gas chromatography (GC). Moreover, the availability of selective and sensitive detectors has made GC the preferred method for quantitative analysis of pesticides. Electron capture detector (ECD) is the common detectors usually employed for the determination of organochlorine (halogen-containing) pesticides, caused by very good response for organochlorine pesticides (Tadeo *et al.*, 2004). Under the optimized condition, the chromatogram obtained for four pesticide standards is shown in Figure 2. All organochlorine pesticides (heptachlor, endosulfan, dieldrin, and *p,p'*-DDT) along with internal standar of aldrin are well separated with good resolution.

In order to validate GC-ECD for simultaneous analysis of these pesticides, some analytical figure of merits namely linearity, precision, sensitivity, and accuracy were determined.

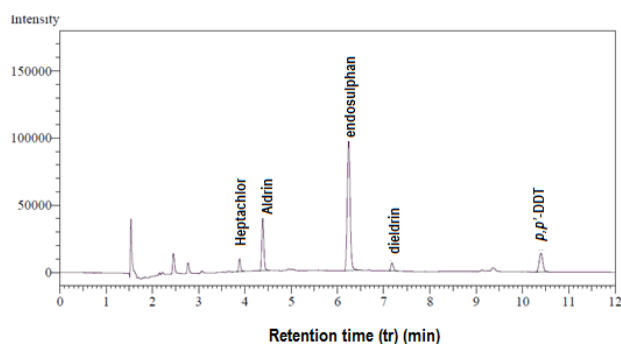


Figure 2. Chromatogram of standard pesticides. Heptachlor (0,02 ng/ μ L), tr = 3,886 min; aldrin (0,1 ng/ μ L); tr = 4,387 min; endosulphan (1,0 ng/ μ L), tr = 6,243 min; dieldrin (0,02 ng/ μ L), tr = 7,180 min; and p,p'-DDT (0,10 ng/ μ L) tr = 10,399 min. Injection volume was set at 1,0 μ L.

3.1. Linearity

A linear regression analysis was carried out by plotting the chromatographic response (chromatogram area) for each organochlorine pesticides (*y*-axis) versus the final concentrations of pesticides (*x*-axis). A set of 10 organochlorine standard solutions in toluene ranging in concentration from 0.005 to 0.08 ppm (heptachlor), 0.25 – 4.00 ppm (endosulphan), 0.005 – 0.08 (dieldrin), and 0.025 – 0.400 (p,p'-DDT) were analyzed in triplicate, and the chromatographic data were used to assess linearity. Regression analysis yielded a coefficient of correlation (*r*) of 0.9989. The *r* values together the regression equation obtained during the linearity assessment were compiled in Table 1. According to Eurachem (1998), the analytical response was linear over certain concentration ranges if *r* obtained is higher than 0.995. Therefore, it can be stated that the used method was linear for analysis of selected organochlorine pesticides at specified linear ranges.

Table 1. The regression equation for the relationship between chromatogram area (*y*-axis) and concentration of pesticides, in ppm (*x*-axis)

| Pesticides | Linear regression equation | Coefficient of correlation (<i>r</i>) |
|------------|----------------------------|-----------------------------------------|
| Heptachlor | $y = 7.3825x + 0.0499$ | 0,9989 |
| Endosulfan | $y = 1.4642x + 1.5833$ | 0,9923 |
| Dieldrin | $y = 7.2959x + 0.0298$ | 0,9995 |
| p,p'-DDT | $y = 3.6035x + 0.0559$ | 0,9981 |

3.2. Sensitivity

Sensitivity of GC with electron capture detector was evaluated by calculating the values of limit of detection (LOD) and limit of quantitation (LOQ). In order to calculate the values of LOD and LOQ, the peak area of ten blank rice samples were measured. LOD and LOQ were calculated as $3.3SD/b$ and $10SD/b$ respectively, where *SD* is the standard deviation of GC chromatogram area and *b* is the slope of calibration curve. When sample blank cannot produce any response, ten independent samples of rice was fortified at the lowest acceptable concentration of each pesticide (Gonzales and Herrador, 2007). The values of LOD obtained were 2.50, 2.97, 1.61, and 2.54 ng/mL for heptachlor, endosulphan, dieldrin, and p,p'-DDT, respectively; meanwhile the LOQ values

obtained are 8.30 (heptachlor), 9.89 (endosulphan), 5.30 (dieldrin) and 8.46 (p,p'-DDT).

3.3. Precision

Precision of analytical method is generally evaluated by calculating relative standard deviation (RSD) or coefficient of variation (CV) of a set of data. Precision of GC method was checked to assess the reproducibility of instrument response to target of analyte. In order to assess the analytical method precision, measurements were done under conditions of repeatability. Repeatability was evaluated by measuring 10 blank rice samples spiked with 0.02 ppm (heptachlor), 1.0 ppm (endosulphan), 0.02 (dieldrin), and 0.01 ppm (p,p'-DDT) under similar conditions (day, analyst, instrument, sample) (ICH, 1994). The RSD values obtained was 4.55, 4.69, 7.85, and 10.90 % for heptachlor, endosulphan, dieldrin, and p,p'-DDT, respectively. According to Horwitz, as cited from Gonzalez and Herrador (2007), the maximum RSD value acceptable for the analyte level of 1 ppm is 16 % (Table 2). Therefore, it can be stated that the developed method exhibited a good precision.

Table 2. The criteria of acceptable RSD values as a function of analyte levels according to Horwitz (Gonzalez and Herrador, 2007)

| Level of analyte | Maximum RSD values |
|------------------|--------------------|
| 100 % | 2.0 |
| 10 % | 2.8 |
| 1 % | 4.0 |
| 0,01 % | 5.7 |
| 100 ppm | 8.0 |
| 10 ppm | 11.3 |
| 1 ppm | 16.0 |
| 100 ppb | 22.6 |
| 10 ppb | 32.0 |
| 1 ppb | 45.3 |

3.3 Accuracy

In this study, the accuracy of analytical method was assessed using standard addition method by calculating the recovery values for each pesticide. These studies were carried out to confirm the lack of analyte losses during sample preparation and matrix interferences during the measurement step (Eurachem, 1998).

Table 3. The recovery percentage of pesticide in fortified rice

| Pesticides | Level of spiked standard (ng/ μ L) | % recovery |
|-------------|----------------------------------------|--------------------|
| Heptachlor | 0.01 | 96.53 \pm 8.65 |
| | 0.02 | 100.25 \pm 2.79 |
| | 0,03 | 95.53 \pm 5.17 |
| Endosulphan | 0.50 | 82.10 \pm 10.35 |
| | 1.00 | 106.50 \pm 3.96 |
| | 1.50 | 109.38 \pm 8.82 |
| Dieldrin | 0.01 | 84.43 \pm 6.45 |
| | 0.02 | 96.78 \pm 2.44 |
| | 0.03 | 110.35 \pm 1.88 |
| p,p'-DDT | 0,05 | 109.25 \pm 4.33 |
| | 0.10 | 111.48 \pm 14.37 |
| | 0.15 | 104.87 \pm 6.19 |

ICH (1994) required that accuracy studies was performed using three different levels of spiked

standards, namely 80 %, 100 %, and 120 % from target of analytes. All analytical steps were performed in three replicates. The recovery percentage values for accuracy studies were shown in Table 3. According to Codex Alimentarius commission (2003), for the level of analyte of > 1 ppm to ≤ 0.01 ppm, the recovery values should be in the range of 60 – 120 %. Therefore, the developed method was accurate for quantification of these pesticides in fortified rice.

4. Conclusion

Gas chromatography with electron capture detection has been successfully developed and validated for simultaneous analysis of four organochlorine pesticides namely heptachlor, endosulphan, dieldrin and p,p-DDT in fortified rice samples. Evaluation of analytical method parameters including linearity, sensitivity, precision and accuracy showed acceptable results. The developed method can also be used for separation and quantitative analysis of rice samples available in market.

5. Acknowledgement

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