Review:

A Review of Marine Bacterial Intracellular and Extracellular Bioactive Compounds as Novel Antibacterial and Anti-Inflammation Agents

Harningsih Karim^{1*}, Arief Azis¹, Ananda Ramadani¹, Anita Anita^{2,3}, Ahyar Ahmad^{4,5}, Hasnah Natsir⁴, Paulina Taba⁴, Suriati Eka Putri⁶, Sarlan Sarlan², Siti Halimah Larekeng^{7,8,9}, and Rizal Irfandi⁶

¹Department of Pharmacy, School of Pharmacy YAMASI, Jl. Mapala 2 Blok D5 No. 10, Makassar 90222, Indonesia

²Doctoral Program, Department of Chemistry, Faculty of Mathematics and Natural Sciences, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia

³Medical Laboratory Technology, Polytechnic Muhammadiyah Makassar, Jl. Dr. Ratulangi No. 101, Makassar 90132, Indonesia

⁴Department of Chemistry, Faculty of Mathematics and Natural Sciences, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia

⁵*Research and Development Center for Biopolymers and Bioproducts, LPPM, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia*

⁶Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Negeri Makassar, Jl. Daeng Tata, Makassar 90244, Indonesia

⁷Faculty of Forestry, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia

⁸Research Collaboration Center for KARST Microbes BRIN-LPPM, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia

⁹KARST Bioprospecting and Society Research Group, Hasanuddin University, Jl. Perintis Kemerdekaan Km. 10, Makassar 90245, Indonesia

* Corresponding author:

email: harningsihkarim@gmail.com

Received: October 23, 2023 Accepted: September 17, 2024

DOI: 10.22146/ijc.90039

Abstract: Unique and varied bioactive compounds produced by the ocean have drawn attention and served as a focus for creating antibacterial and anti-inflammatory agents. As part of the approach for locating these research sources, databases such as PubMed, Science Direct, MDPI, Google Scholar, Springer Link, Web of Science, Scopus, and Wiley Online Library were used to identify completed studies. Numerous intriguing bioactive compounds have so far been isolated from marine bacteria. A crucial resource in the ongoing search for novel peptides, proteins, lipids, nucleosides, enzymes, alkaloids, polyketides, and terpenoids is the diversity of marine bacterium strains. This review summarizes several bacterial intracellular and extracellular bioactive compounds that have been applied as antibacterial and anti-inflammatory agents in 2016–2024, which we present in the form of structures, species sources, and evaluations of these compounds' antibacterial and anti-inflammatory agents for these compounds is the future for utilizing biomaterials from marine bacteria that are promising in the future for industrial-scale production of antibacterial and anti-inflammatory agents.

Keywords: antibacterial; anti-inflammation; bioactive compounds; marine bacteria

INTRODUCTION

The word "antibiotic" derives from the Greek word "antibiosis", which means "against life" [1-2]. In the past, people used to think of antibiotics as chemical substances produced by one type of bacteria but poisonous to another [2]. Based on this description, antibiotics were initially generally defined as compounds produced by a bacterium or other organisms [3] that at low quantities could stop the growth of or kill other bacteria [4]. However, in recent times, this definition has been expanded to encompass antimicrobial substances manufactured partially or entirely synthetically. Some antibiotics can only prevent other bacteria from growing, while others can destroy them [5]. Bactericidal and bacteriostatic substances prevent bacteria growth [6]. Antibiotics are classified as antiviral, antifungal, and antibacterial to correspond with the types of microbes they antagonize, even though they are commonly called "antibacterial" [7-8].

Most antibacterial medications used in medicine are made from natural materials or closely resemble them [2,9]. A total of 98 small-scale commercial combinations of antibacterial medications with identifiable structural motifs derived from nature were present between 1981 and 2005 [10]. Selection necessitated by bacteria leads to random chromosomal changes, which allow the bacteria to "adapt" to life in the presence of particular antibacterial chemicals [11-12]. Resistance to bactericidal drugs develops faster because of the rapid expansion of the genetic material encoding adherence [13]. The emergence of methicillin-resistant *Staphylococcus aureus* (MRSA), vancomycin-resistant *Enterococcus* (VRE), and epidemiclevel *Pseudomonas* resistant to fluoroquinolones in 2010 marked the turning point of bacterial infections [14-15].

Since the majority of antibiotic classes were discovered between 1940 and 1960, a period known as the "golden age" of antibiotic research [16], it is believed that the chemical arsenal available for treating human microbiological infections is sufficient. The development of germ resistance to antibiotics has, however, been fast. Microbes are subjected to selective pressure by antimicrobial agents, which causes random selection and chromosomal alterations that increase their susceptibility to environmental factors such as antibiotics, disinfectants, and chemical compounds [17-19]. Resistance-producing genetic material quickly replicates and spreads [20]. Additionally, antimicrobial drug persistence is greatly accelerated by the excessive and inappropriate use of antimicrobial medicines [21-22]. Antibiotic-resistant bacteria are categorized as agents that are detrimental to human health by the World Health Organization (WHO) and the Centers for Disease Control and Prevention (CDC) [23-24]. These organizations also predict that by 2050, antimicrobial resistance might be responsible for up to 100 trillion US dollars of global economic losses or 10 million fatalities annually [25].

The marine environment covers 70% of the planet's surface. It has a much greater phylogenetic diversity than the various types of bacteria that have evolved to withstand particular environmental conditions (such as low light, high pressure, high acidity, low temperature, and varying oxygen content and salinity), which lead to the production of primary and secondary metabolites with unique structures and complex bioactivity [26-28]. Bioactive compounds produced by microorganisms, particularly bacteria, are thought to be interesting due to their structural variety and potential [29-30]. Numerous new pharmaceutical compounds can be made from sediments and bacteria produced by marine species [31]. Related marine bacteria secrete biologically active substances used as nutrients (such as carbon) from the host to defend themselves from potentially dangerous environmental components [32]. This review covered recent and trending topics such as intracellular and extracellular bioactive compounds with antibacterial and antiinflammatory activity from marine bacteria.

METHODS USED FOR LITERATURE COLLECTION

Online databases like PubMed, Science Direct, MDPI, Google Scholar, Springer Link, Web of Science, Scopus, Research Gate, and Wiley Online Library were used to conduct the literature search for this review. In addition, the selection of articles used exclusion and inclusion criteria using articles written in English. We also followed several steps recommended by PRISMA, such as writing a structured summary of each article, assessing studies and research methods, identifying the limitations of each study, drawing conclusions, and looking for implications of the main findings. Reviews are arranged based on the order of topics. This paper reviews the latest developments in marine bacterial intracellular and extracellular bioactive compounds as antibacterial and anti-inflammatory agents. The search approach is centered on principal keywords that are utilized in different combinations, such as bioactive compounds of marine bacteria, antibacterial and anti-inflammatory

DIFFERENT MARINE BACTERIA PHYLA PRODUCE ANTIMICROBIAL AND ANTI-INFLAMMATION AGENT

marine bacteria, and the structure of bioactive marine

bacteria [33].

Marine environments, particularly the ocean, are rich sources of diverse microorganisms, including bacteria with unique metabolic capabilities. Marine bacteria have evolved to produce bioactive compounds as a survival strategy against competing microorganisms or as a defense mechanism against predators. These compounds can exhibit antimicrobial and anti-inflammatory properties, making them potentially valuable for therapeutic and pharmaceutical applications [34-35]. These marine bacteria phyla have evolved unique biosynthetic pathways to produce a wide range of bioactive compounds, many exhibiting antimicrobial and anti-inflammatory properties [36-39]. Some marine bacterial phyla generate antibacterial and anti-inflammatory including the mechanism of action of every compound, as shown in Table 1. These compounds can potentially be developed into new therapeutic agents or serve as lead compounds for drug discovery efforts.

INTRACELLULAR BIOACTIVE COMPOUNDS OF MARINE BACTERIA

Peptides

The class of proteins known as antimicrobial peptides exhibits broad-spectrum antimicrobial action and is well-known to be effective against various infections. Numerous antimicrobial peptides (AMPs) exhibit an immediate and direct antibacterial effect by disrupting the physical integrity of the microbial membrane and/or by crossing the membrane to reach the cytoplasm of bacteria and act on intracellular targets [39]. These chemicals partially resolved the microbial resistance conundrum, which restricted the use of many effective antimicrobial medicines [52]. AMPs are a new treatment option for cancer patients since they also have antibacterial and anticancer capabilities. AMPs are currently mostly used in medicine to treat inflammation, wound healing, and infections caused by harmful microorganisms [53-54]. Despite these promising findings, only a few studies have focused peptide sources on exophytic bacteria derived from marine biota. In recent years, other studies have also reported using various proteins and/or peptides from marine microorganisms as antibacterial and anticancer agents [55-56]. Our group has shown that protein hydrolysates obtained from different sources of microsymbiont marine algae could effectively inhibit pathogenic bacteria and HeLa

Phyla	Marine bacteria species	Mechanism of action	Ref.
Proteobacteria	Pseudoalteromonas sp.	Antimicrobial and antitumor compounds: proteins, polypeptides,	[40-41]
		and pigments	
	<i>Vibrio</i> sp.	Antimicrobial and anti-inflammatory compounds: android and	[42-43]
		brindle	
Actinobacteria	Salinispora sp.	Antimicrobial and anticancer compounds: salinosporamide A and	[44-45]
		salinosporamide K	
	Streptomyces sp.	Antimicrobial, anticancer, antiviral compounds: lantimycins and	[46]
		streptonigrin	
Firmicutes	Bacillus sp.	Antimicrobial compounds: surfactants, iturins and fengycin	[47-49]
Bacteroidetes	Flavobacterium sp.	Antimicrobial compounds: flavocyclomycin	[50]
Cyanobacteria	Lyngba sp., Oscillatoria sp.	Antimicrobial and anti-inflammatory compounds: dolastatin,	[51]
		curacin, and lyngbyatoxin	

Table 1. Marine bacterial phyla generate antibacterial and anti-inflammatory compounds

cancer cell lines [57-58]. These findings collectively indicate the potential of intracellular bioactive compounds of aquatic bacteria and peptide-based approaches in combating various diseases, including cancer, antibacterial, and anti-inflammatory. Further research could lead to the development of new and effective therapeutic agents for these health conditions. The structure of antibacterial peptide compounds from marine bacteria based on the results of previous studies is summarized in Table 2.

Lipid

The antimicrobial lipid classes that have been studied the most over the years include sterols, fatty acids, diacylglycerols (DAG), monoacylglycerols (MAG), and terpene derivatives [65]. The efficacy of these lipids against different microorganisms is dependent on both the medium's pH and their chemical makeup [66]. The structure-activity relationship between free fatty acids (non-esterified) and bacteria is dependent on the acyl chain length, stereochemistry, degree of unsaturation, and esterified form of the fatty acids [67]. In Grampositive bacteria, long-chain fatty acids (C12 or above) are effective at low concentrations and under pH dependence [68-69], but short-chain fatty acids (C6 or below) are effective at high concentrations and pH dependence [70]. It has also been observed that unsaturated isomers are more potent against Gramnegative bacteria and that cis-isomers are more active than trans-isomers [71]. Table 3 displays the composition of the antibacterial lipid molecules produced by various marine bacteria.

Peptide Compounds	Structure	Marine bacteria species	Mechanism of action	Ref.
Diketopiperazine: cyclo-(L-valiyl-D- proline)		<i>R. japonica</i> strain KMM 9513	Effective against methicillin- resistant <i>S. aureus</i> Antitumor, antiviral	[59]
cyclo-(L-phenylalanyl- D-proline)		S. Tocher	Anteancer	[00]
Tetrapeptides: cyclo-(Leu-Pro-Ile- Pro)	6 ^m 0 1 ^m 1 ^m 1 ^m 0 6 ^m 0	B. amyloliquefaciens GAS 00152	Antitumor, antibacterial	[61]
cyclo-(Tyr-Pro-Phe- Gly)	4/ 4/ 4/ 4/ 4/ 4/ 4/ 4/ 4/ 4/	<i>Ruegeria</i> sp.	Effective against <i>S. aureus</i> ATCC 25923, <i>E. coli</i> ATCC 25922	[62]
	6' HN 1" 1" 3"' 5"'		Antitumor, antihelminth	[63]
Desotamide		Streptomyces sp. NRRL 21611, Streptomyces sp. JAMM992, S. scopuliridis SCSIO Z	Effective against <i>S. aureus</i> , MRSE pneumoniae, <i>Mycobacterium</i> sp.	[64]

Table 2. Structure of antibacterial peptide compounds from marine bacteria



Table 3. Structure of antibacterial lipid compounds from marine bacteria

Nucleosides

Marine microorganisms are capable of producing a variety of nucleosides with unusual structures and biological characteristics [72]. The discovery of these remarkable biological characteristics in marine nucleosides has sparked a great deal of research into the synthesis of their various analogs and the ongoing evaluation of their biological factors, such as analgesic, anti-inflammatory, antibacterial, and anticancer activities [73]. The search for novel analogs of natural nucleosides with potential biological properties has accelerated the field of nucleoside chemistry research [74]. Based on earlier research findings, the composition and structure of antibacterial nucleoside compounds in various marine bacteria are outlined in Table 4.

Enzymes

by The marine enzymes generated microorganisms have numerous applications in the bioindustry. Utilizing microorganisms with elevated yields and production rates is crucial in manufacturing industrial enzymes. Enzymes such as amylase, casein, lipase, gelatinase, and DNase are found in microbes isolated from severe maritime environments. These enzymes are thermally stable, resistant to a wide range of pH values and other harsh conditions required for industrial applications and may have biological applications related to human health [80]. According to the research findings, enzymes produced by marine bacteria with a score of 4 or higher were selected for 16S rRNA molecular analysis. About 161 bacterial isolates

Nucleoside compounds	Structure	Marine bacteria	Mechanism of action	Ref.
Rocheicoside A		S. rochei 06CM016	Effective against <i>E. faecium</i> DSM 13590: vancomycin resistant; <i>E. coli</i> O157:H7 RSKK 234: streptomycin, sulfisoxazole and tetracycline-resistant; <i>S. aureus</i> : MRSA; <i>P. aureginosa</i> ATCC 27853	[78]
Formycin A		N. interforma, S. kaniharaensis SF- 557	Antiviral and antitumor activities	[79]
Pyrazofurin A	HO Formycin A (FOR-A) H ₂ N HO HO Pyrazofurin A (PRF-A)	S. candidus NRRL 3601, S. lavendulae	Antitumor activities	[79- 80]

Table 4. Structure of antibacterial nucleoside compounds from marine bacteria

produced 68.7% of amylase, 88.3% of lipase, and 68.7% of protease in their secretions. Phylogenetic analysis led to the discovery of 4 major phyla: actinobacteria, proteobacteria, firmicutes, and bacteroidetes [81-82]. Several types of antibacterial enzymes contained in several marine bacteria are shown in Table 5.

EXTRACELLULAR BIOACTIVE COMPOUNDS OF MARINE BACTERIA

Alkaloid

Natural products are small molecules that are isolated from biological sources. They have long been recognized for their tremendous promise in human medicine, and in recent years, their popularity has continued to expand [9]. In the field of pharmaceutical development, natural product discovery has overtaken (combinatorial) chemistry thanks to the advent of novel technologies like more precise analytical methods or enhanced genome mining algorithms [84]. The most common antibacterial substances in marine environments are alkaloids [85]. A class of compounds known as marine alkaloids has shown promise in medicine [86]. Table 6 lists a few kinds of marine bacteria with a particular antibacterial alkaloid.

Polyketides

One of the most important problems in contemporary biotechnology is still discovering new antibiotics, as dangerous microorganisms quickly become resistant to them [91]. The latter leads to a growing number of incurable or difficult-to-treat bacterial infections, which may eventually be among the leading causes of death. This is why it is so important for modern medicine to discover novel antibacterial compounds [92]. Polyketide-based natural antimicrobial marine products are a diverse class with various structural modifications, including antiviral, antibacterial, anticancer, and other effects [93]. Table 7 summarizes the composition of antibacterial polyketide chemicals in different marine bacteria based on findings from earlier research.

Enzyme compounds	Structure	Marine bacteria	Mechanism of action	Ref.
L-Asparaginase		<i>Streptomyces</i> sp.	Effective against MRSA, <i>E. coli</i> , and tumor therapy	[81- 82]
Amylase		V. alginolyticus	Effective against Gram-positive bacteria	[83]
Protease		V. harveyi, V. costicola	Effective against MRSA, <i>E. coli, and anticancer</i>	[83]

Alkaloid compounds	Structure	Marine bacteria	Mechanism of action	Ref.
Marinopyrrole A	R_1 H_N Cl_0 R_2 Cl_0 R_2 Cl_0 R_2	Streptomyces sp.	Effective against MRSA	[87]
Bacilsubteramide A		B. subterraneus 11593, Micromonospora sp. A258	Effective against MRSA and Gram- positive bacteria, anti-allergic bioactivities	[88]

Table 6. Structure of antibacterial alkaloid compounds from marine bacteria

289



Table 7. Structure of antibacterial polyketides compounds from marine bacteria

Polyketides compounds	Structure	Marine bacteria	Mechanism of action	Ref.
N-acetyl-N- demethylmayamycin	OH O OH R NH	<i>Streptomyces</i> sp. 182SMLY	Effective against MRSA, induced apoptosis in the glioma cells	[94]
Streptophenazines		<i>Streptomyces</i> sp. 182SMLY	Effective against MRSA	[95]
Actinomycins D	$\begin{array}{c} \cdot \\ - \\ - \\ N \\ - \\ - \\ N \\ - \\ - \\ - \\ -$	<i>Streptomyces</i> sp. ZZ338	Effective against MRSA, <i>E. coli</i> , inhibiting the proliferation of glioma cells	[96]
Salinosporamide A		S. tropica	Potential anticancer, antimalarial, antibacterial MRSA, <i>M. tuberculosis</i> , and anti-inflammatory properties	[44,97]



Terpenoid

Numerous organisms that produce bioactive natural chemicals are part of the vast biological diversity seen in the marine environment [99]. Many different structural groups and biological roles of chemicals can be found in marine microorganisms [43]. Natural compounds with antibacterial properties are influenced by a variety of biotic and abiotic factors found in the marine environment, including temperature, nutrition, salinity, interactions with other microbes, and other factors [100]. Marine bacteria use terpenoids as important metabolites [101]. Table 8 displays the antibacterial terpenoids structural composition in marine microorganisms.

ANTIBACTERIAL ACTIVITY OF MARINE BACTERIA

Marine environments are rich reservoirs of diverse microorganisms, a vast and largely unexplored source of microbial diversity, including bacteria that have evolved unique metabolic capabilities and biosynthetic pathway [106]. These marine bacteria have demonstrated remarkable potential for producing bioactive compounds with antibacterial properties, offering promising avenues for combating bacterial infections, particularly those caused by multidrug-resistant pathogen [43]. Marine bacteria produce a wide range of bioactive compounds with antibacterial properties [107]. These compounds have attracted significant attention due to their potential applications in combating bacterial infections, particularly those caused by drug-resistant pathogens [108]. The following are important details regarding marine bacteria's antimicrobial activity. First is diverse sources. Antibacterial compounds-producing marine bacteria have been isolated from sediments, saltwater, marine invertebrates (such as sponges, corals, and tunicates), and algae [109-110]. Second, diversity in structure. The antibacterial substances generated by marine bacteria have a noteworthy diversity in structure, encompassing peptides, polyketides, terpenoids, alkaloids, and additional categories of natural products [111]. Third, mechanisms of action. Marine bacterial antibacterial

Terpenoids	Structure	Marine	Mechanism of action	Ref.
compounds		bacteria		
Napyradiomycin		S. aculeolatus	Effective against methicillin-resistant <i>S. aureus</i> , exhibited moderate cytotoxicity against four human cancer cell lines SF-268	[102]
Dixiamycin A	но 20 н	Streptomyces	Effective against herpes simplex virus-1 (HSV-1) in vitro	[103]
	HO H ₃ C CH ₃	sp.	S. aureus, Acinetobacter sp., and MRSA	

Table 8. Structure of antibacterial terpenoid compounds from marine bacteria



compounds can act through various mechanisms, such as disrupting cell membranes, inhibiting cell wall synthesis, interfering with protein synthesis, or targeting essential enzymes and cellular processes in bacterial cells [30,112-113]. Fourth, broad-spectrum antibacterial activity. Numerous marine bacterial compounds have been shown to be effective against Gram-positive and Gram-negative bacteria, including those resistant to several drugs [114]. Fifth, novel chemical scaffolds. Marine bacterial metabolites are appealing prospects for the creation of novel antibacterial drugs because they frequently have distinct chemical structures and scaffolds that differ from those of conventional antibiotics [108]. Sixth, biofilm inhibition. Some marine bacterial compounds have been identified to suppress the production of biofilms, which is a major cause of antibiotic resistance and persistent infections. These compounds also have antibacterial qualities [19,115].

The composition of the cell wall is a major factor in determining how susceptible or resistant bacteria are to various antimicrobial agents [116]. It is noteworthy that several antimicrobial chemicals derived from marine bacteria possess broad-spectrum activity, which enables them to target both Gram-positive and Gram-negative bacteria efficiently [30]. This wide range of activity can be explained by their capacity to target conserved cellular structures or processes, such as those that interfere with vital enzymes, disturb membrane integrity, or impede protein synthesis, which is shared by both species of bacteria [117]. Bacteria are often classified as Gram-positive or Gram-negative depending on the unique components of their cell walls [113]. Unlike Gram-positive bacteria, which have a thick peptidoglycan layer encircling their cytoplasmic membrane, Gram-negative bacteria have an outer membrane and a thin peptidoglycan layer covering their cytoplasmic membrane [118-121].

Cell wall permeability is a crucial factor in target intracellular inhibition, and these discrepancies have wider implications for antibacterial efficacy [122]. The peptidoglycan coating of Gram-positive bacteria [123] allows antibacterial substances to pass through it efficiently (e.g., *S. aureus* and *E. faecium*). Many times, the chemical characteristics necessary for piercing the glycolipid layer are highly distinct and peculiar to the membranes of Gram-negative bacteria (e.g., *E. coli* and *P. aeruginosa*) [124-125]. In reality, the development of specialized Gram-positive antibiotics has been successful, while the development of Gram-negative antibiotics has not [126-127]. The structure of the Gram-positive cell wall depicts the thick peptidoglycan layer, which also contains wall-associated teichoic acids and membrane-associated lipoteichoic acids, as shown in Fig. 1 [127]. In addition, the structure of the Gram-negative cell wall includes the peptidoglycan layer, periplasmic space, and outer membrane. The external leaflet of the outer membrane is predominately lipopolysaccharides. The outer membrane and peptidoglycan are linked together by Braun's lipoprotein, as shown in Fig. 2 [128].

The antibacterial activity of substances made by marine bacteria is frequently assessed using a variety of techniques, such as the following. The first method is disk diffusion assay. In this method, sterile disks containing the marine bacterial extract or purified compound are placed on an agar plate and inoculated with the test bacterial strain. After incubation, the zone of inhibition around the disk is measured, indicating the compound's ability to inhibit bacterial growth [129]. The second method is a well diffusion assay. It is a good diffusion assay and comparable to the disk diffusion assay, but it uses wells punched into the agar plate to disseminate the marine bacterial extract or chemical compound that is dispensed into the wells [130-131]. The third method is minimum inhibitory concentration (MIC) determination. This method involves preparing serial dilutions of the marine bacterial extract or compound in a liquid growth medium, followed by inoculation with the test bacterial strain. The MIC is defined as the lowest concentration of the compound that inhibits visible bacterial growth after incubation [121]. The fourth method is minimum bactericidal concentration (MBC) determination. This method is comparable to that of determining MIC in that it involves plating the contents of the MIC wells onto fresh agar plates and looking for signs of bacterial growth. The MBC is the lowest concentration of the compound that kills the bacterial [132].

Time-kill assays technique entails subjecting the test bacterial strain to various marine bacterial chemical concentrations for a predetermined amount of time. Colony-forming units (CFUs) are counted and plated to assess the amount of viable bacterial cells in samples that are taken at regular intervals. Understanding the kinetics of the antibacterial action is possible using this method [133]. Biofilm inhibition and eradication tests assess



Fig 1. The Gram-positive cell wall structure depicts the thick peptidoglycan layer, which also contains wall-associated teichoic acids and membrane-associated lipoteichoic acids [127]



Fig 2. The Gram-negative cell wall structure includes the peptidoglycan layer, periplasmic space, and outer membrane. The external leaflet of the outer membrane is predominately lipopolysaccharides. Braun's lipoprotein links the outer membrane and peptidoglycan together [128]

whether marine bacterial chemicals can disrupt and break up pre-existing biofilms or prevent new ones from forming. It is possible to use a variety of quantitative and qualitative techniques, including microscopic procedures, metabolic activity tests, and crystal violet staining [133].

The purpose of combination studies is to ascertain whether the marine bacteria component, when coupled with other antibiotics or antimicrobial agents, demonstrates synergistic antibacterial action [133]. Numerous biochemical, molecular, and microscopic techniques can be used to shed light on the specific mechanisms by which the chemicals found in marine exert their antibacterial bacteria effects. These mechanisms may include the disruption of membranes, the inhibition of cell wall formation, or interference with protein synthesis [134]. It is significant to remember that these techniques can be adjusted and improved in accordance with the particular needs of the experiment, the characteristics of the marine bacterial compound, and the target species or strain of bacteria. Furthermore, in vivo investigations employing suitable models might be conducted to assess the antibacterial effectiveness and toxicity of a promising marine bacterial agent.

ANTI-INFLAMMATORY ACTIVITIES OF MARINE BACTERIA

Marine bacteria frequently have a substantial and varied supply of biologically active compounds with suspected anti-inflammatory properties, contributing to the exploration of novel therapeutic agents for various inflammatory conditions [92,111]. These microorganisms showed promising anti-inflammatory properties through different pathways [43]. In vitro and in vivo models of inflammation, such as cell-based assays, animal models of inflammatory disorders, and clinical investigations, have all demonstrated the antiinflammatory properties of marine bacterial metabolites [135]. These bioactive substances have demonstrated possible beneficial effects in the treatment of inflammatory diseases and disorders such as rheumatoid arthritis, asthma, inflammatory bowel diseases, and neurodegenerative disorders [136]. Although these bacteria have evolved to survive in unique and frequently unpleasant settings, they have produced a diverse range of secondary metabolites with a variety of biological actions, including the ability to reduce inflammation [137]. These marine bacterial metabolites' distinct chemical structures and modes of action present exciting prospects for the creation of novel anti-inflammatory medications and treatment approaches [43]. One significant anti-inflammatory activity of marine bacteria is the inhibition of pro-inflammatory mediators [136]. Studies have identified marine bacteria-derived compounds that can modulate cytokine production, such as tumor necrosis factor (TNF)-alpha and interleukins, play crucial roles in which the inflammatory response [138]. Marine bacteria help to lessen inflammation and related tissue damage by preventing the release of these pro-inflammatory chemicals [139].

In many disorders, oxidative stress and inflammation are significant factors. An immune defense process known as inflammation is triggered by noxious stimuli such as mechanical trauma, burns, microbial infections, allergies, and other noxious stimuli [140-141]. The recommended carrageenan-induced paw edema method was used to measure the antiinflammatory activity [140,142-143]. Marine microorganisms are highly favored in drug development research due to their potential for producing bioactive compounds. Antimycin-type depsipeptides (Fig. 3), namely urauchimycin D (1) and somalimycin (2), were isolated from a mutant strain of Streptomyces somaliensis SCSIO ZH66. These compounds were found to suppress the production of interleukin-5 in splenocytes induced by ovalbumin in mice. Among the depsipeptides, compound 1 exhibited strong inhibitory activity with an IC₅₀ value of 0.57 µM, while compound 2 showed milder effects (> 10μ M). Additionally, these depsipeptides displayed minimal cytotoxicity against human umbilical vein endothelial cells, with LD₅₀ values of 62.6, 34.6, and 192.9 µM, respectively. The (+)- and (-)-actinoxocine (3a, 3b) were isolated from a marinederived Streptomyces sp. and showed inhibition on TNFa protein release in LPS- and Pam3CSK4-induced RAW 264.7 mouse macrophages, respectively [144]. The



Fig 3. Structures of anti-inflammatory compounds from marine bacteria

macrolide caniferolide A (4) from *S. caniferus* could block NF κ Bp65 translocation to the nucleus and showed inhibition on the production of pro-inflammatory cytokines (IL-1 β , IL-6, and TNF α), the release of NO, and the activities of iNOS, JNK, and p38 in LPS induced BV2 microglial cells [145].

Honaucin A (5), isolated from a marine-polyketides cyanobacterium, has been found to inhibit mouse ear edema [146]. The crude pigment extracts of marine *Brevibacterium* sp. were an effective compound for antiinflammatory activity using carrageenan-induced paw edema on Wistar male rats [38]. Cyclomarins (6) was isolated from a marine *Streptomyces* sp. have been found to have interesting lead structures for developing drugs against tuberculosis and malaria [147]. These findings demonstrate the potential of marine-derived compounds as promising candidates for developing anti-inflammatory agents, which could have significant implications for treating various inflammatory conditions and diseases.

DISCUSSION

Marine environments are the source of all life, and it is thought that this "primordial soup" is where the earliest forms of life first emerged [148]. With its varied bacteria, the marine environment evolves specific adaptation mechanisms due to thriving in a different type of climate [127,149]. These mechanisms may be helpful for their defense, and the outcomes of these adaptations may benefit humans in various ways [150]. One such marine bacterial defense strategy against predators is the production of bioactive metabolites [151]. This means that marine bacteria are probably a very promising source for new antibiotic and anti-inflammation manufacturers [147-150]. Considering their distinct living environments and metabolic processes, these investigations indicate that marine bacteria have great potential for creating novel physiologically active substances [151]. The fight against the threat posed by the rise in infections caused by antibiotic-resistant organisms includes the hunt for new antibiotics and anti-inflammation as a key component. A key strategy in the fight against the threat posed by the growth in diseases brought on by drug-resistant microbes is the search for novel antibiotics and anti-inflammation [152]. According to literature reviews, most antibiotics and anti-inflammations used today are synthesized from relatively few scaffold molecules. Antimicrobial and antiinflammation drugs for harmful bacteria are undoubtedly becoming more prevalent. However, the rate of finding and creating new, potent antibacterial and antiinflammation substances is slowing down.

CONCLUSION

Marine bacteria are important providers of structurally diverse and distinctive intracellular such as peptides, lipids, nucleosides, enzymes, and extracellular bioactive compounds such as alkaloids, polyketides, and terpenoids. We highlighted the diversity of marine bacteria, which has helped researchers identify novel antibacterial and anti-inflammatory agents and supports the notion that research holds great promise for developing biomaterials in the future for large-scale industrial production of antibacterial and antiinflammatory agent applications.

ACKNOWLEDGMENTS

The authors thank DSTI Hasanuddin University and the Department of Pharmacy, School of Pharmacy YAMASI, Makassar, Indonesia. Grant vocation partially supported this work, PKDN scheme with contract No. I.2.1.082.113/IV/2024.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

The study was designed by Harningsih Karim, Ahyar Ahmad, and Anita. Ahyar Ahmad, Arief Azis, Ananda Ramadani, and Harningsih Karim collected the data. Ahyar Ahmad and Hasnah Natsir conducted the data analysis and interpretation, while Harningsih Karim, Anita, and Paulina Taba wrote the manuscript. Anita, Suriati Eka Putri, and Sarlan prepared figure/table. Ahyar Ahmad, Hasnah Natsir, Siti Halimah Larekeng, and Paulina Taba revised the manuscript. All authors discussed the results and contributed to the final manuscript.

REFERENCES

- Nehra, S., Gothwal, R.K., Varshney, A.K., Solanki, P.S., Chandra, S., Meena, P., Trivedi, P.C., and Ghosh, P., 2021, "Chapter 19 - Bio-Management of *Fusarium* spp. Associated with Fruit Crops" in *Fungi Bio-Prospects in Sustainable Agriculture, Environment and Nano-Technology*, Eds. Sharma, V.K., Shah, M.P., Parmar, S., and Kumar, A., Academic Press, Cambridge, MA, US, 475–505.
- Hutchings, M.I., Truman, A.W., and Wilkinson, B., 2019, Antibiotics: Past, present and future, *Curr. Opin. Microbiol.*, 51, 72–80.
- Purssell, E., 2019, "Antimicrobials" in Understanding Pharmacology in Nursing Practice, Eds. Hood, P., and Khan, E., Springer International Publishing, Cham, Switzerland, 147–165.

Harningsih Karim et al.

- [4] Doolin, T., Gross, S., and Siryaporn, A., 2020,
 "Physical Mechanisms of Bacterial Killing by Histones" in *Physical Microbiology*, Eds. Duménil, G., and van Teeffelen, S., Springer International Publishing, Cham, Switzerland, 117–133.
- [5] Hasan, T.H., and Al-Harmoosh, R.A., 2020, Mechanisms of antibiotics resistance in bacteria, *Syst. Rev. Pharm.*, 11 (6), 817–823.
- [6] Gomes, C.F., Gomes, J.H., and da Silva, E.F., 2020, Bacteriostatic and bactericidal clays: An overview, *Environ. Geochem. Health*, 42 (11), 3507–3527.
- Brochot, A., Guilbot, A., Haddioui, L., and Roques,
 C., 2017, Antibacterial, antifungal, and antiviral effects of three essential oil blends, *MicrobiologyOpen*, 6 (4), e00459.
- [8] Senerovic, L., Opsenica, D., Moric, I., Aleksic, I., Spasić, M., and Vasiljevic, B., 2020, "Quinolines and Quinolones as Antibacterial, Antifungal, Antivirulence, Antiviral and Anti-parasitic Agents" in Advances in Microbiology, Infectious Diseases and Public Health: Volume 14, Eds. Donelli, G., Springer International Publishing, Cham, Switzerland, 37–69.
- [9] Atanasov, A.G., Zotchev, S.B., Dirsch, V.M., The International Natural Product Sciences Taskforce, and Supuran, C.T., 2021, Natural products in drug discovery: Advances and opportunities, *Nat. Rev. Drug Discovery*, 20 (3), 200–216.
- [10] Young, R.J., Flitsch, S.L., Grigalunas, M., Leeson, P.D., Quinn, R.J., Turner, N.J., and Waldmann, H., 2022, The time and place for nature in drug discovery, *JACS Au*, 2 (11), 2400–2416.
- [11] Hassan, A.Y., Lin, J.T., Ricker, N., and Anany, H., 2021, The age of phage: Friend or foe in the new dawn of therapeutic and biocontrol applications, *Pharmaceuticals*, 14 (3), 199.
- [12] Jian, Z., Zeng, L., Xu, T., Sun, S., Yan, S., Yang, L., Huang, Y., Jia, J., and Dou, T., 2021, Antibiotic resistance genes in bacteria: Occurrence, spread, and control, *J. Basic Microbiol.*, 61 (12), 1049–1070.
- [13] Zhang, F., and Cheng, W., 2022, The mechanism of bacterial resistance and potential bacteriostatic strategies, *Antibiotics*, 11 (9), 1215.

- [14] Turner, N.A., Sharma-Kuinkel, B.K., Maskarinec, S.A., Eichenberger, E.M., Shah, P.P., Carugati, M., Holland, T.L., and Fowler, V.G., 2019, Methicillinresistant *Staphylococcus aureus*: An overview of basic and clinical research, *Nat. Rev. Microbiol.*, 17 (4), 203–218.
- [15] Munita, J.M., and Arias, C.A., 2016, "Mechanisms of Antibiotic Resistance" in *Virulence Mechanisms* of Bacterial Pathogens, Eds. Kudva, I.T., Cornick, N.A., Plummer, P.J., Zhang, Q., Nicholson, T.L., Bannantine, J.P., and Bellaire, B.H., ASM Press, Washington, DC, US, 481–511.
- [16] Ribeiro da Cunha, B., Fonseca, L.P., and Calado, C.R.C., 2019, Antibiotic discovery: Where have we come from, where do we go?, *Antibiotics*, 8 (2), 45.
- [17] Kannappan, A., Sivaranjani, M., Srinivasan, R., Rathna, J., Pandian, S.K., and Ravi, A.V., 2017, Inhibitory efficacy of geraniol on biofilm formation and development of adaptive resistance in *Staphylococcus epidermidis* RP62A, *J. Med. Microbiol.*, 66 (10), 1506–1515.
- [18] Li, Z., Zhang, L., Song, Q., Wang, G., Yang, W., Tang, H., Srinivasan, R., Lin, L., and Lin, X., 2021, Proteomics analysis reveals bacterial antibiotics resistance mechanism mediated by *ahslyA* against enoxacin in *Aeromonas hydrophila*, *Front. Microbiol.*, 12, 699415.
- [19] Srinivasan, R., Santhakumari, S., Poonguzhali, P., Geetha, M., Dyavaiah, M., and Xiangmin, L., 2021, Bacterial biofilm inhibition: A focused review on recent therapeutic strategies for combating the biofilm mediated infections, *Front. Microbiol.*, 12 (5), 676458.
- [20] Li, Z., Wang, Y., Li, X., Lin, Z., Lin, Y., Srinivasan, R., and Lin, X., 2019, The characteristics of antibiotic resistance and phenotypes in 29 outermembrane protein mutant strains in *Aeromonas hydrophila*, *Environ*. *Microbiol.*, 21 (12), 4614– 4628.
- [21] Podolsky, S.H., 2018, The evolving response to antibiotic resistance (1945–2018), *Palgrave Commun.*, 4 (1), 124.

- [22] Alexpandi, R., Prasanth, M.I., Ravi, A.V., Balamurugan, K., Durgadevi, R., Srinivasan, R., De Mesquita, J.F., and Pandian, S.T.K., 2019, Protective effect of neglected plant *Diplocyclos palmatus* on quorum sensing mediated infection of Serratia marcescens and UV-A induced photoaging in model *Caenorhabditis elegans, J. Photochem. Photobiol., B*, 201, 111637.
- [23] World Health Organization, 2024, WHO Updates List of Drug-Resistant Bacteria Most Threatening to Human Health, https://www.who.int/news/item/17-05-2024-who-updates-list-of-drug-resistantbacteria-most-threatening-to-human-health.
- [24] De Oliveira, D.M.P., Forde, B.M., Kidd, T.J., Harris, P.N.A., Schembri, M.A., Beatson, S.A., Paterson, D.L., and Walker, M.J., 2020, Antimicrobial resistance in ESKAPE pathogens, *Clin. Microbiol. Rev.*, 33 (3), e00181-19.
- [25] O'Neill, J., 2016, Tackling Drug-Resistant Infections Globally: Final Report and Recommendations, The Review on Antimicrobial Resistance, 1–84.
- [26] Johnson, S.M., and Watson, J.R., 2021, Novel environmental conditions due to climate change in the world's largest marine protected areas, *One Earth*, 4 (11), 1625–1634.
- [27] Sunagawa, S., Acinas, S.G., Bork, P., Bowler, C., Acinas, S.G., Babin, M., Bork, P., Boss, E., Bowler, C., Cochrane, G., de Vargas, C., and Tara Oceans Coordinators, 2020, *Tara* Oceans: Towards global ocean ecosystems biology, *Nat. Rev. Microbiol.*, 18 (8), 428–445.
- [28] Choudhary, A., Naughton, L.M., Montánchez, I., Dobson, A.D.W., and Rai, D.K., 2017, Current status and future prospects of marine natural products (MNPs) as antimicrobials, *Mar. Drugs*, 15 (9), 272.
- [29] Pham, J.V., Yilma, M.A., Feliz, A., Majid, M.T., Maffetone, N., Walker, J.R., Kim, E., Cho, H.J., Reynolds, J.M., Song, M.C., Park, S.R., and Yoon, Y.J., 2019, A review of the microbial production of bioactive natural products and biologics, *Front. Microbiol.*, 10, 1404.
- [30] Srinivasan, R., Kannappan, A., Shi, C., and Lin, X., 2021, Marine bacterial secondary metabolites: A

treasure house for structurally unique and effective antimicrobial compounds, *Mar. Drugs*, 19 (10), 530.

- [31] Jagannathan, S.V., Manemann, E.M., Rowe, S.E., Callender, M.C., and Soto, W., 2021, Marine actinomycetes, new sources of biotechnological products, *Mar. Drugs*, 19 (7), 365.
- [32] Dang, N.P., Landfald, B., and Willassen, N.P., 2016, Biological surface-active compounds from marine bacteria, *Environ. Technol.*, 37 (9), 1151–1158.
- [33] Barzkar, N., Sukhikh, S., and Babich, O., 2024, Study of marine microorganism metabolites: New resources for bioactive natural products, *Front. Microbiol.*, 14, 1285902.
- [34] Stincone, P., and Brandelli, A., 2020, Marine bacteria as source of antimicrobial compounds, *Crit. Rev. Biotechnol.*, 40 (3), 306–319.
- [35] Tabarzad, M., Atabaki, V., and Hosseinabadi, T., 2020, Anti-inflammatory activity of bioactive compounds from microalgae and cyanobacteria by focusing on the mechanisms of action, *Mol. Biol. Rep.*, 47 (8), 6193–6205.
- [36] Núñez-Montero, K., and Barrientos, L., 2018, Advances in Antarctic research for antimicrobial discovery: A comprehensive narrative review of bacteria from Antarctic environments as potential sources of novel antibiotic compounds against human pathogens and microorganisms of industrial importance, *Antibiotics*, 7 (4), 90.
- [37] Quintero, M., Velásquez, A., Jutinico, L.M., Jiménez-Vergara, E., Blandón, L.M., Martinez, K., Lee, H.S., and Gómez-León, J., 2018, Bioprospecting from marine coastal sediments of Colombian Caribbean: Screening and study of antimicrobial activity, *J. Appl. Microbiol.*, 125 (3), 753–765.
- [38] Srilekha, V., Krishna, G., Seshasrinivas, V., and Charya, M.A.S., 2017, Antibacterial and antiinflammatory activities of marine *Brevibacterium* sp., *Res. Pharm. Sci.*, 12 (4), 283–289.
- [39] Yasir, M., 2018, Analysis of bacterial communities and characterization of antimicrobial strains from cave microbiota, *Braz. J. Microbiol.*, 49 (2), 248– 257.

- [40] Bosi, E., Fondi, M., Orlandini, V., Perrin, E., Maida, I., de Pascale, D., Tutino, M.L., Parrilli, E., Lo Giudice, A., Filloux, A., and Fani, R., 2017, The pangenome of (Antarctic) *Pseudoalteromonas* bacteria: Evolutionary and functional insights, *BMC Genomics*, 18 (1), 93.
- [41] Offret, C., Desriac, F., Le Chevalier, P., Mounier, J., Jégou, C., and Fleury, Y., 2016, Spotlight on antimicrobial metabolites from the marine bacteria *Pseudoalteromonas*: Chemodiversity and ecological significance, *Mar. Drugs*, 14 (7), 1-26.
- [42] Buijs, Y., Isbrandt, T., Zhang, S.D., Larsen, T.O., and Gram, L., 2020, The Antibiotic andrimid produced by *Vibrio corallilyticus* increases expression of biosynthetic gene clusters and antibiotic production in *Photobacterium galatheae*, *Front. Microbiol.*, 11, 622055.
- [43] Karthikeyan, A., Joseph, A., and Nair, B.G., 2022, Promising bioactive compounds from the marine environment and their potential effects on various diseases, *J. Genet. Eng. Biotechnol.*, 20 (1), 14.
- [44] McCauley, E.P., Piña, I.C., Thompson, A.D., Bashir, K., Weinberg, M., Kurz, S.L., and Crews, P., 2020, Highlights of marine natural products having parallel scaffolds found from marine-derived bacteria, sponges, and tunicates, *J. Antibiot.*, 73 (8), 504–525.
- [45] Kim, H., Kim, S., Kim, M., Lee, C., Yang, I., and Nam, S.J., 2020, Bioactive natural products from the genus *Salinospora*: A review, *Arch. Pharmacal Res.*, 43 (12), 1230–1258.
- [46] Anteneh, Y.S., Yang, Q., and Brown, M.H., 2021, Antimicrobial activities of marine sponge-associated bacteria, *Microorganisms*, 9 (1), 171.
- [47] Kubicki, S., Bollinger, A., Katzke, N., Jaeger, K.E., Loeschcke, A., and Thies, S., 2019, Marine biosurfactants: Biosynthesis, structural diversity and biotechnological applications, *Mar. Drugs*, 17 (7), 408.
- [48] Gudiña, E.J., Teixeira, J.A., and Rodrigues, L.R., 2016, Biosurfactants produced by marine microorganisms with therapeutic applications, *Mar. Drugs*, 14 (2), 38.
- [49] Xiao, S., Chen, N., Chai, Z., Zhou, M., Xiao, C., Zhao, S., and Yang, X., 2022, Secondary metabolites from

marine-derived *Bacillus*: A comprehensive review of origins, structures, and bioactivities, *Mar. Drugs*, 20 (9), 567.

- [50] Hamidi, M., Kozani, P.S., Kozani, P.S., Pierre, G., Michaud, P., and Delattre, C., 2020, Marine bacteria versus microalgae: Who is the best for biotechnological production of bioactive compounds with antioxidant properties and other biological applications?, *Mar. Drugs*, 18 (1), 28.
- [51] Khalifa, S.A.M., Shedid, E.S., Saied, E.M., Jassbi, A.R., Jamebozorgi, F.H., Rateb, M.E., Du, M., Abdel-Daim, M.M., Kai, G.Y., Al-Hammady, M.A.M., Xiao, J., Guo, Z., and El-Seedi, H.R., 2021, Cyanobacteria—From the oceans to the potential biotechnological and biomedical applications, *Mar. Drugs*, 19 (5), 241.
- [52] Ponnappan, N., Budagavi, D.P., and Chugh, A., 2017, CyLoP-1: Membrane-active peptide with cellpenetrating and antimicrobial properties, *Biochim. Biophys. Acta, Biomembr.*, 1859 (2), 167–176.
- [53] Costa, F., Teixeira, C., Gomes, P., and Martins, M.C.L., 2019, "Clinical Application of AMPs" in Antimicrobial Peptides: Basics for Clinical Application, Springer Singapore, Singapore, 281– 298.
- [54] Mahlapuu, M., Håkansson, J., Ringstad, L., and Björn, C., 2016, Antimicrobial peptides: An emerging category of therapeutic agents, *Front. Cell. Infect. Microbiol.*, 6, 194.
- [55] Ahmad, A., Asmi, N., Karim, H., Massi, M.N., Wahid, I., and Sugrani, A., 2020, Characterization of anti-dengue and cytotoxic activity of protein hydrolysates from the exophytic bacteria of brown algae Sargassum sp., J. Appl. Pharm. Sci., 11 (02), 039–045.
- [56] Macedo, M.W.F.S., da Cunha, N.B., Carneiro, J.S.A., da Costa, R.A., de Alencar, S.A., Cardoso, M.H., Franco, O.L., and Dias, S.C., 2021, Marine organisms as a rich source of biologically active peptides, *Front. Mar. Sci.*, 8, 667764.
- [57] Asmi, N., Ahmad, A., Karim, H., Massi, M.N., Natsir, H., Karim, A., Taba, P., Dwyana, Z., and Ibrahim, M., 2020, Antibacterial effect of protein

and protein hydrolysates isolated from bacteria *Enterobacter hormaechei* associated with marine algae *Sargassum* sp., *Rasayan J. Chem.*, 13 (3), 1606–1611.

- [58] Sugrani, A., Ahmad, A., Djide, M.N., and Natsir, H., 2020, Two novel antimicrobial and anti-cancer peptides prediction from *Vibrio* sp. strain ES25, *J. Appl. Pharm. Sci.*, 10 (08), 058–066.
- [59] Kalinovskaya, N.I., Romanenko, L.A., and Kalinovsky, A.I., 2017, Antibacterial low-molecularweight compounds produced by the marine bacterium *Rheinheimera japonica* KMM 9513(T), *Antonie van Leeuwenhoek*, 110 (5), 719–726.
- [60] Al-Rawahi, A.N., Abed, R.M.M., Rehman, N.U., Rafiq, K., Khan, A., Khan, A.L., Khan, M., Halim, S.A., Al-Senafi, F., Mahmoud, H., and Al-Harrasi, A., 2023, New sulfur-containing diketopiperazine from marine-derived bacteria *Streptomyces rochei* sp. 81 with potent carbonic anhydrase II inhibition, *Chem. Nat. Compd.*, 59 (2), 346–350.
- [61] Chakraborty, S., Tai, D.F., Lin, Y.C., and Chiou, T.W., 2016, Antitumor and antimicrobial activity of some cyclic tetrapeptides and tripeptides derived from marine bacteria, *Mar. Drugs*, 13 (5), 3029–3045.
- [62] Dahiya, R., Kumar, S., Khokra, S.L., Gupta, S.V., Sutariya, V.B., Bhatia, D., Sharma, A., Singh, S., and Maharaj, S., 2018, Toward the synthesis and improved biopotential of an *N*-methylated analog of a proline-rich cyclic tetrapeptide from marine bacteria, *Mar. Drugs*, 16 (9), 305.
- [63] Fazal, A., Webb, M.E., and Seipke, R.F., 2020, The desotamide family of antibiotics, *Antibiotics*, 9 (8), 452.
- [64] Alves, E., Dias, M., Lopes, D., Almeida, A., Domingues, M.D., and Rey, F., 2020, Antimicrobial lipids from plants and marine organisms: An overview of the current state-of-the-art and future prospects, *Antibiotics*, 9 (8), 441.
- [65] de Carvalho, C.C.C.R., and Caramujo, M.J., 2018, The various roles of fatty acids, *Molecules*, 23 (10), 2583.
- [66] Alexandri, E., Ahmed, R., Siddiqui, H., Choudhary, M.I., Tsiafoulis, C.G., and Gerothanassis, I.P., 2017, High resolution NMR spectroscopy as a structural

and analytical tool for unsaturated lipids in solution, *Molecules*, 22 (10), 1663.

- [67] Fischer, C.L., 2020, Antimicrobial activity of hostderived lipids, *Antibiotics*, 9 (2), 75.
- [68] Yoon, B.K., Jackman, J.A., Valle-González, E.R., and Cho, N.J., 2018, Antibacterial free fatty acids and monoglycerides: Biological activities, experimental testing, and therapeutic applications, *Int. J. Mol. Sci.*, 19 (4), 1114.
- [69] Machado, M.G., Sencio, V., and Trottein, F., 2021, Short-chain fatty acids as a potential treatment for infections: A closer look at the lungs, *Infect. Immun.*, 89 (9), e0018821.
- [70] Eberlein, C., Baumgarten, T., Starke, S., and Heipieper, H.J., 2018, Immediate response mechanisms of Gram-negative solvent-tolerant bacteria to cope with environmental stress: *Cistrans* isomerization of unsaturated fatty acids and outer membrane vesicle secretion, *Appl. Microbiol. Biotechnol.*, 102 (6), 2583–2593.
- [71] Rotter, A., Barbier, M., Bertoni, F., Bones, A.M., Cancela, M.L., Carlsson, J., Carvalho, M.F., Cegłowska, M., Chirivella-Martorell, J., Conk Dalay, M., Cueto, M., Dailianis, T., Deniz, I., Díaz-Marrero, A.R., Drakulovic, D., Dubnika, A., Edwards, C., Einarsson, H., Erdoğan, A., Eroldoğan, O.T., Ezra, D., Fazi, S., FitzGerald, R.J., Gargan, L.M., Gaudêncio, S.P., Gligora Udovič, M., Ivošević DeNardis, N., Jónsdóttir, R., Kataržytė, M., Klun, K., Kotta, J., Ktari, L., Ljubešić, Z., Lukić Bilela, L., Mandalakis, M., Massa-Gallucci, A., Matijošytė, I., Mazur-Marzec, H., Mehiri, M., Nielsen, S.L., Novoveská, L., Overlingė, D., Perale, G., Ramasamy, P., Rebours, C., Reinsch, T., Reyes, F., Rinkevich, B., Robbens, J., Röttinger, E., Rudovica, V., Sabotič, J., Safarik, I., Talve, S., Tasdemir, D., Theodotou Schneider, X., Thomas, O.P., Toruńska-Sitarz, A., Varese, G.C., and Vasquez, M.I., 2021, The essentials of marine biotechnology, Front. Mar. Sci., 8, 629629.
- [72] Hussain, F., Rahman, F.I., Saha, P., Mikami, A., Osawa, T., Obika, S., and Abdur Rahman, S.M., 2022, Synthesis of sugar and nucleoside analogs and

evaluation of their anti-cancer and analgesic potentials, *Molecules*, 27 (11), 3499.

- [73] Yates, M.K., and Seley-Radtke, K.L., 2019, The evolution of anti-viral nucleoside analogues: A review for chemists and non-chemists. Part II: Complex modifications to the nucleoside scaffold, *Antiviral Res.*, 162, 5–21.
- [74] Mani, P., Dineshkumar, G., Jayaseelan, T., Deepalakshmi, K., Ganesh Kumar, C., and Senthil Balan, S., 2016, Antimicrobial activities of a promising glycolipid biosurfactant from a novel marine *Staphylococcus saprophyticus* SBPS 15, 3 *Biotech*, 6 (2), 163.
- [75] Wei, T., Zhao, C., Quareshy, M., Wu, N., Huang, S., Zhao, Y., Yang, P., Mao, D., and Chen, Y., 2021, A glycolipid glycosyltransferase with broad substrate specificity from the marine bacterium "*Candidatus; Pelagibacter* sp." strain HTCC7211, *Appl. Environ. Microbiol.*, 87 (14), e00326-21.
- [76] Jenssen, M., Kristoffersen, V., Motiram-Corral, K., Isaksson, J., Rämä, T., Andersen, J.H., Hansen, E.H., and Hansen, K.Ø., 2021, Chlovalicin B, a chlorinated sesquiterpene isolated from the marine mushroom *Digitatispora marina*, *Molecules*, 26 (24), 7560.
- [77] Aksoy, S.Ç., Uzel, A., and Bedir, E., 2016, Cytosinetype nucleosides from marine-derived *Streptomyces rochei* 06CM016, *J. Antibiot.*, 69 (1), 51–56.
- [78] Zhang, M., Zhang, P., Xu, G., Zhou, W., Gao, Y., Gong, R., Cai, Y.S., Cong, H., Deng, Z., Price, N.P.J., Mao, X., and Chen, W., 2020, Comparative investigation into formycin A and pyrazofurin A biosynthesis reveals branch pathways for the construction of C-Nucleoside scaffolds, *Appl. Environ. Microbiol.*, 86 (2), e01971-19.
- [79] Xu, G., Kong, L., Gong, R., Xu, L., Gao, Y., Jiang, M., Cai, Y.S., Hong, K., Hu, Y., Liu, P., Deng, Z., Price, N.P.J., and Chen, W., 2018, Coordinated biosynthesis of the purine nucleoside antibiotics aristeromycin and coformycin in actinomycetes, *Appl. Environ. Microbiol.*, 84 (22), e01860-18.
- [80] Kamala, K., and Sivaperumal, P., 2017, Biomedical applications of enzymes from marine actinobacteria, *Adv. Food Nutr. Res.*, 80, 107–123.

- [81] Izadpanah Qeshmi, F., Homaei, A., Fernandes, P., and Javadpour, S., 2018, Marine microbial Lasparaginase: Biochemistry, molecular approaches and applications in tumor therapy and in food industry, *Microbiol. Res.*, 208, 99–112.
- [82] Bunpa, S., Sermwittayawong, N., and Vuddhakul, V., 2016, Extracellular enzymes produced by *Vibrio* alginolyticus isolated from environments and diseased aquatic animals, *Procedia Chem.*, 18, 12– 17.
- [83] Gaudêncio, S.P., Bayram, E., Lukić Bilela, L., Cueto, M., Díaz-Marrero, A.R., Haznedaroglu, B.Z., Jimenez, C., Mandalakis, M., Pereira, F., Reyes, F., and Tasdemir, D., 2023, Advanced methods for natural products discovery: Bioactivity screening, dereplication, metabolomics profiling, genomic sequencing, databases and informatic tools, and structure elucidation, *Mar. Drugs*, 21 (5), 308.
- [84] Almeida, M.C., Resende, D.I.S.P., da Costa, P.M., Pinto, M.M.M., and Sousa, E., 2021, Tryptophan derived natural marine alkaloids and synthetic derivatives as promising antimicrobial agents, *Eur. J. Med. Chem.*, 209, 112945.
- [85] Lu, W.Y., Li, H.J., Li, Q.Y., and Wu, Y.C., 2021, Application of marine natural products in drug research, *Bioorg. Med. Chem.*, 35, 116058.
- [86] Kemung, H.M., Tan, L.T.H., Khan, T.M., Chan, K.G., Pusparajah, P., Goh, B.H., and Lee, L.H., 2018, *Streptomyces* as a prominent resource of future anti-MRSA drugs, *Front. Microbiol.*, 9, 2221.
- [87] Xie, C.L., Xia, J.M., Su, R.Q., Li, J., Liu, Y., Yang, X.W., and Yang, Q., 2018, Bacilsubteramide A, a new indole alkaloid, from the deep-sea-derived *Bacillus subterraneus* 11593, *Nat. Prod. Res.*, 32 (21), 2553–2557.
- [88] Newaz, A.W., Yong, K., Lian, X.Y., and Zhang, Z., 2022, Streptoindoles A–D, novel antimicrobial indole alkaloids from the marine-associated actinomycete *Streptomyces* sp. ZZ1118, *Tetrahedron*, 104, 132598.
- [89] Anjum, K., Kaleem, S., Yi, W., Zheng, G., Lian, X., and Zhang, Z., 2019, Novel antimicrobial indolepyrazines A and B from the marine-

associated *Acinetobacter* sp. ZZ1275, *Mar. Drugs*, 17 (2), 89.

- [90] Thabit, A.K., Crandon, J.L., and Nicolau, D.P., 2015, Antimicrobial resistance: Impact on clinical and economic outcomes and the need for new antimicrobials, *Expert Opin. Pharmacother.*, 16 (2), 159–177.
- [91] Uddin, T.M., Chakraborty, A.J., Khusro, A., Zidan, B.M.R.M., Mitra, S., Bin Emran, T., Dhama, K., Ripon, M.K.H., Gajdács, M., Sahibzada, M.U.K., Hossain, M.J., and Koirala, N., 2021, Antibiotic resistance in microbes: History, mechanisms, therapeutic strategies and future prospects, *J. Infect. Public Health*, 14 (12), 1750–1766.
- [92] Huang, C., Yang, C., Zhu, Y., Zhang, W., Yuan, C., and Zhang, C., 2018, Marine bacterial aromatic polyketides from host-dependent heterologous expression and fungal mode of cyclization, *Front. Chem.*, 6, 528.
- [93] Liang, Y., Xie, X., Chen, L., Yan, S., Ye, X., Anjum, K., Huang, H., Lian, X., and Zhang, Z., 2016, Bioactive polycyclic quinones from marine *Streptomyces* sp. 182SMLY, *Mar. Drugs*, 14 (1), 10.
- [94] Liang, Y., Chen, L., Ye, X., Anjum, K., Lian, X.Y., and Zhang, Z., 2017, New streptophenazines from marine *Streptomyces* sp. 182SMLY, *Nat. Prod. Res.*, 31 (4), 411–417.
- [95] Zhang, X., Ye, X., Chai, W., Lian, X.Y., and Zhang, Z., 2016, New metabolites and bioactive actinomycins from marine-derived *Streptomyces* sp. ZZ338, *Mar. Drugs*, 14 (10), 181.
- [96] Francis, A., and Chakraborty, K., 2021, Marine macroalga-associated heterotroph *Bacillus velezensis* as prospective therapeutic agent, *Arch. Microbiol.*, 203 (4), 1671–1682.
- [97] Chakraborty, K., Kizhakkekalam, V.K., and Joy, M., 2022, Polyketide-derived macrobrevins from marine macroalga-associated *Bacillus amyloliquefaciens* as promising antibacterial agents against pathogens causing nosocomial infections, *Phytochemistry*, 193, 112983.
- [98] Subramani, R., and Sipkema, D., 2019, Marine rare actinomycetes: A promising source of structurally

diverse and unique novel natural products, Mar. Drugs, 17 (5), 249.

- [99] Gozari, M., Alborz, M., El-Seedi, H.R., and Jassbi, A.R., 2021, Chemistry, biosynthesis and biological activity of terpenoids and meroterpenoids in bacteria and fungi isolated from different marine habitats, *Eur. J. Med. Chem.*, 210, 112957.
- [100] Núñez-Pons, L., Shilling, A., Verde, C., Baker, B.J., and Giordano, D., 2020, Marine terpenoids from polar latitudes and their potential applications in biotechnology, *Mar. Drugs*, 18 (8), 401.
- [101] Pereira, F., Almeida, J.R., Paulino, M., Grilo, I.R., Macedo, H., Cunha, I., Sobral, R.G., Vasconcelos, V., and Gaudêncio, S.P., 2020, Antifouling napyradiomycins from marine-derived actinomycetes *Streptomyces aculeolatus*, *Mar. Drugs*, 18 (1), 63.
- [102] Jiang, Y.C., Feng, H., Lin, Y.C., and Guo, X.R., 2016, New strategies against drug resistance to herpes simplex virus, *Int. J. Oral Sci.*, 8 (1), 1–6.
- [103] Velmurugan, P., Venil, C.K., Veera Ravi, A., and Dufossé, L., 2020, Marine bacteria is the cell factory to produce bioactive pigments: A prospective pigment source in the ocean, *Front. Sustainable Food Syst.*, 4, 589655.
- [104] Podilapu, A.R., Emmadi, M., and Kulkarni, S.S., 2018, Expeditious synthesis of ieodoglucomides A and B from the marine-derived bacterium *Bacillus licheniformis*, *Eur. J. Org. Chem.*, 2018 (24), 3230– 3235.
- [105]Shu, W.S., and Huang, L.N., 2022, Microbial diversity in extreme environments, *Nat. Rev. Microbiol.*, 20 (4), 219–235.
- [106] Chinnathambi, A., Salmen, S.H., Al-Garadi, M.A., Wainwright, M., and Ali Alharbi, S., 2023, Marine actinomycetes: An endless source of potentially therapeutic novel secondary metabolites and other bioactive compounds, *J. King Saud Univ.*, Sci., 35 (9), 102931.
- [107] Mba, I.E., and Nweze, E.I., 2022, Antimicrobial peptides therapy: An emerging alternative for treating drug-resistant bacteria, *Yale J. Biol. Med.*, 95 (4), 445–463.

- [108] Zha, X., Ji, R., and Zhou, S., 2024, Marine bacteria: A source of novel bioactive natural products, *Curr. Med. Chem.*, 31 (41), 6842–6854.
- [109] Ameen, F., AlNadhari, S., and Al-Homaidan, A.A., 2021, Marine microorganisms as an untapped source of bioactive compounds, *Saudi J. Biol. Sci.*, 28 (1), 224–231.
- [110] Wang, P., Huang, X., Jiang, C., Yang, R., Wu, J., Liu, Y., Feng, S., and Wang, T., 2024, Antibacterial properties of natural products from marine fungi reported between 2012 and 2023: A review, *Arch. Pharmacal Res.*, 47 (6), 505–537.
- [111] Sarkar, P., Yarlagadda, V., Ghosh, C., and Haldar, J., 2017, A review on cell wall synthesis inhibitors with an emphasis on glycopeptide antibiotics, *MedChemComm*, 8 (3), 516–533.
- [112] Navarro, P.P., Vettiger, A., Ananda, V.Y., Llopis, P.M., Allolio, C., Bernhardt, T.G., and Chao, L.H., 2022, Cell wall synthesis and remodelling dynamics determine division site architecture and cell shape in *Escherichia coli, Nat. Microbiol.*, 7 (10), 1621–1634.
- [113] Liu, Y., Shi, J., Tong, Z., Jia, Y., Yang, K., and Wang, Z., 2020, Potent broad-spectrum antibacterial activity of amphiphilic peptides against multidrugresistant bacteria, *Microorganisms*, 8 (9), 1398.
- [114] Varela, M.F., Stephen, J., Lekshmi, M., Ojha, M., Wenzel, N., Sanford, L.M., Hernandez, A.J., Parvathi, A., and Kumar, S.H., 2021, Bacterial resistance to antimicrobial agents, *Antibiotics*, 10 (5), 593.
- [115] Zhang, Q.Y., Yan, Z.B., Meng, Y.M., Hong, X.Y., Shao, G., Ma, J.J., Cheng, X.R., Liu, J., Kang, J., and Fu, C.Y., 2021, Antimicrobial peptides: Mechanism of action, activity and clinical potential, *Mil. Med. Res.*, 8 (1), 48.
- [116] Lin, T.Y., and Weibel, D.B., 2016, Organization and function of anionic phospholipids in bacteria, *Appl. Microbiol. Biotechnol.*, 100 (10), 4255–4267.
- [117] Dörr, T., Moynihan, P.J., and Mayer, C., 2019, Editorial: Bacterial cell wall structure and dynamics, *Front. Microbiol.*, 10, 2051.
- [118] Fivenson, E.M., Rohs, P.D.A., Vettiger, A., Sardis, M.F., Torres, G., Forchoh, A., and Bernhardt, T.G.,

2023, A role for the Gram-negative outer membrane in bacterial shape determination, *Proc. Natl. Acad. Sci. U. S. A.*, 120 (35), e2301987120.

- [119]Zgurskaya, H.I., and Rybenkov, V.V., 2020, Permeability barriers of Gram-negative pathogens, *Ann. N. Y. Acad. Sci.*, 1459 (1), 5–18.
- [120] Borisova, M., Gaupp, R., Duckworth, A., Schneider, A., Dalügge, D., Mühleck, M., Deubel, D., Unsleber, S., Yu, W., Muth, G., Bischoff, M., Götz, F., and Mayer, C., 2016, Peptidoglycan recycling in Gram-positive bacteria is crucial for survival in stationary phase, *MBio*, 7 (5), e00923-16.
- [121] Sun, J., Rutherford, S.T., Silhavy, T.J., and Huang, K.C., 2022, Physical properties of the bacterial outer membrane, *Nat. Rev. Microbiol.*, 20 (4), 236– 248.
- [122] Sperandeo, P., Martorana, A.M., and Polissi, A., 2017, Lipopolysaccharide biogenesis and transport at the outer membrane of Gram-negative bacteria, *Biochim. Biophys. Acta, Mol. Cell Biol. Lipids*, 1862 (11), 1451–1460.
- [123] Tavares, T.D., Antunes, J.C., Padrão, J., Ribeiro, A.I., Zille, A., Amorim, M.T.P., Ferreira, F., and Felgueiras, H.P., 2020, Activity of specialized biomolecules against Gram-positive and Gramnegative bacteria, *Antibiotics*, 9 (6), 314.
- [124] Kloska, A., Cech, G.M., Sadowska, M., Krause, K., Szalewska-Pałasz, A., and Olszewski, P., 2020, Adaptation of the marine bacterium *Shewanella baltica* to low temperature stress, *Int. J. Mol. Sci.*, 21 (12), 4338.
- [125] Miller, S.I., and Salama, N.R., 2018, The Gramnegative bacterial periplasm: Size matters, *PLoS Biol.*, 16 (1), e2004935.
- [126] Gefen, O., Chekol, B., Strahilevitz, J., and Balaban, N.Q., 2017, TD test: Easy detection of bacterial tolerance and persistence in clinical isolates by a modified disk-diffusion assay, *Sci. Rep.*, 7 (1), 41284.
- [127] Rütten, A., Kirchner, T., and Musiol-Kroll, E.M., 2022, Overview on strategies and assays for antibiotic discovery, *Pharmaceuticals*, 15 (10), 1302.

- [128] Weinstein, M.P., and Lewis, J.S., 2020, The clinical and laboratory standards institute subcommittee on antimicrobial susceptibility testing: Background, organization, functions, and processes, *J. Clin. Microbiol.*, 58 (3), e01864-19.
- [129] Rodríguez-Melcón, C., Alonso-Calleja, C., García-Fernández, C., Carballo, J., and Capita, R., 2021, Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) for twelve antimicrobials (biocides and antibiotics) in eight strains of *Listeria monocytogenes*, *Biology*, 11 (1), 46.
- [130] Balouiri, M., Sadiki, M., and Ibnsouda, S.K., 2016, Methods for *in vitro* evaluating antimicrobial activity: A review, *J. Pharm. Anal.*, 6 (2), 71–79.
- [131] Hudson, M.A., and Lockless, S.W., 2022, Elucidating the mechanisms of action of antimicrobial agents, *MBio*, 13 (3), e02240-21.
- [132] Patil, K.R., Mahajan, U.B., Unger, B.S., Goyal, S.N., Belemkar, S., Surana, S.J., Ojha, S., and Patil, C.R., 2019, Animal models of inflammation for screening of anti-inflammatory drugs: Implications for the discovery and development of phytopharmaceuticals, *Int. J. Mol. Sci.*, 20 (18), 4367.
- [133] Elbandy, M., 2022, Anti-inflammatory effects of marine bioactive compounds and their potential as functional food ingredients in the prevention and treatment of neuroinflammatory disorders, *Molecules*, 28 (1), 2.
- [134] Petersen, L.E., Kellermann, M.Y., and Schupp, P.J., 2020, "Secondary Metabolites of Marine Microbes: From Natural Products Chemistry to Chemical Ecology" in YOUMARES 9 The Oceans: Our Research, Our Future: Proceedings of the 2018 conference for YOUng MArine RESearcher in Oldenburg, Germany, Springer International Publishing, Cham, Switzerland, 159–180.
- [135] Florean, C., Dicato, M., and Diederich, M., 2022, Immune-modulating and anti-inflammatory marine compounds against cancer, *Semin. Cancer Biol.*, 80, 58–72.
- [136] Parolini, C., 2024, The role of marine n-3 polyunsaturated fatty acids in inflammatory-based

disease: The case of rheumatoid arthritis, Mar. Drugs, 22 (1), 17.

- [137] Baral, P., Udit, S., and Chiu, I.M., 2019, Pain and immunity: Implications for host defence, *Nat. Rev. Immunol.*, 19 (7), 433–447.
- [138]Zhang, S., Chen, Y., Zhu, J., Lu, Q., Cryle, M.J., Zhang, Y., and Yan, F., 2023, Structural diversity, biosynthesis, and biological functions of lipopeptides from *Streptomyces*, *Nat. Prod. Rep.*, 40 (3), 557–594.
- [139] Karim, N., Khan, I., Khan, W., Khan, I., Khan, A., Halim, S.A., Khan, H., Hussain, J., and Al-Harrasi, A., 2019, Anti-nociceptive and anti-inflammatory activities of asparacosin A involve selective cyclooxygenase 2 and inflammatory cytokines inhibition: An *in-vitro*, *in-vivo*, and *in-silico* approach, *Front. Immunol.*, 10, 581.
- [140] Menzel, A., Samouda, H., Dohet, F., Loap, S., Ellulu, M.S., and Bohn, T., 2021, Common and novel markers for measuring inflammation and oxidative stress *ex vivo* in research and clinical practice-which to use regarding disease outcomes, *Antioxidants*, 10 (3), 414.
- [141] Li, H., Huang, H., Hou, L., Ju, J., and Li, W., 2017, Discovery of Antimycin-type depsipeptides from a *wbl* gene mutant strain of deepsea-derived *Streptomyces somaliensis* SCSIO ZH66 and their effects on pro-inflammatory cytokine production, *Front. Microbiol.*, 8, 678.
- [142] Alvariño, R., Alonso, E., Lacret, R., Oves-Costales, D., Genilloud, O., Reyes, F., Alfonso, A., and Botana, L.M., 2019, Caniferolide A, a macrolide from *Streptomyces caniferus*, attenuates neuroinflammation, oxidative stress, amyloidbeta, and tau pathology *in vitro*, *Mol. Pharmaceutics*, 16 (4), 1456–1466.
- [143] Mascuch, S.J., Boudreau, P.D., Carland, T.M., Pierce, N.T., Olson, J., Hensler, M.E., Choi, H., Campanale, J., Hamdoun, A., Nizet, V., Gerwick, W.H., Gaasterland, T., and Gerwick, L., 2018, Marine natural product honaucin A attenuates inflammation by activating the Nrf2-ARE pathway, J. Nat. Prod., 81 (3), 506–514.

- [144] Kazmaier, U., and Junk, L., 2021, Recent developments on the synthesis and bioactivity of ilamycins/rufomycins and cyclomarins, marine cyclopeptides that demonstrate anti-malaria and anti-tuberculosis activity, *Mar. Drugs*, 19 (8), 446.
- [145] Kumar, P.S., 2021, "Chapter One Introduction to Marine Biology" in *Modern Treatment Strategies for Marine Pollution*, Elsevier, Cambridge, MA, US, 1–10.
- [146] Overmann, J., and Lepleux, C., 2016, "Marine Bacteria and Archaea: Diversity, Adaptations, and Culturability" in *The Marine Microbiome: An* Untapped Source of Biodiversity and Biotechnological Potential, Eds. Stal, L.J., and Cretoiu, M.S., Springer International Publishing, Cham, Switzerland, 21–55.
- [147] Johnson, L.A., and Hug, L.A., 2019, Distribution of reactive oxygen species defense mechanisms across domain bacteria, *Free Radical Biol. Med.*, 140, 93– 102.
- [148] Joseph, A., 2017, "Chapter 9 Oceans: Abode of

Nutraceuticals, Pharmaceuticals, and Biotoxins" in *Investigating Seafloors and Oceans*, Elsevier, Cambridge, MA, US, 493–554.

- [149] Wiese, J., and Imhoff, J.F., 2019, Marine bacteria and fungi as promising source for new antibiotics, *Drug Dev. Res.*, 80 (1), 24–27.
- [150] Cardoso, J., Nakayama, D.G., Sousa, E., and Pinto,
 E., 2020, Marine-derived compounds and prospects for their antifungal application, *Molecules*, 25 (24), 5856.
- [151] Chinemerem N.D., Ugwu, M.C., Oliseloke A.C., Al-Ouqaili, M.T.S., Chinedu I.J., Victor C.U., and Saki, M., 2022, Antibiotic resistance: The challenges and some emerging strategies for tackling a global menace, J. Clin. Lab. Anal., 36 (9), e24655.
- [152] Ahmed, S.K., Hussein, S., Qurbani, K., Ibrahim, R.H., Fareeq, A., Mahmood, K.A., and Mohamed, G.M., 2024, Antimicrobial resistance: Impacts, challenges, and future prospects, *J. Med., Surg., Public Health*, 2, 100081.